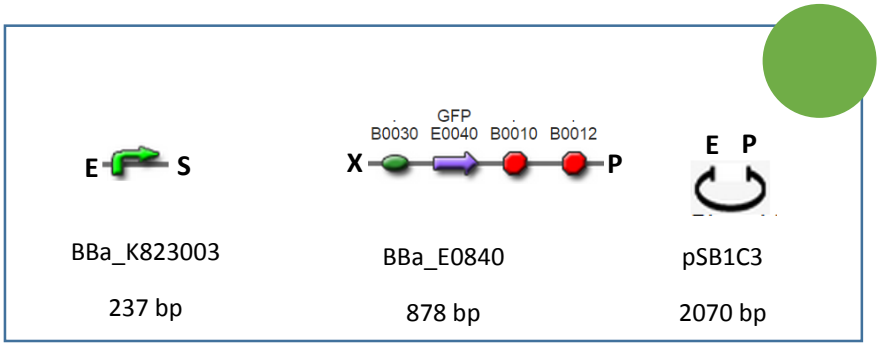
Assembly:

K I



1st Day

EXSP Digestion (see **Enzymatic Digestion Protocol**)

Parts	ng/ul	Volume to 2,5 ug (ul)	Buffer x10 (ul)	EcoRI (ul)	XbaI (ul)	SpeI (ul)	PstI (ul)	H_2O to 50ul (ul)
BBa_K823003	151,4	17,0	5	1	-	1	-	26
BBa_E0840	156,8	15,9	5	-	1	-	1	27,1
pSB1C3	107,3	24,3	5	1	-	-	1	20,7

Repeat this digestion only if you run out of stock

2nd Day

Gel Purification

- See **Kit Wizard SV gel and PCR clean up Promega Protocol**
- Quantify digestion products

Parts	ng/ul	260/280
BBa_K823003 (ES)	13,4	1,85
BBa_E0840 (XP)	16,6	1,79
pSB1C3 (EP)	24,3	2,83

Obs: 260/280 in a quality parameter that tells you if your sample is contaminated with proteins. The greater it is compared to 1 the less contaminants you have.

Ligation (see **Ligation Protocol**)

Linear Plasmid 50 ng	2 ul	
Insert : Plasmid 5:1 (BBa_K823003) ; 3:1 (BBa_E0840)	BBa_K823003	BBa_E0840
	3 ul	4 ul
10x T4 DNA Buffer	2 ul	
T4 DNA ligase 1u	1 ul	
H_20 to 20 ul	8 ul	

Obs: To determinate the amount of DNA necessary we used the following equation

$$Insert\ ng = plasmid\ ng \times \frac{insert\ bp}{plasmid\ bp} \times insert{:}\,plasmid\ ratio$$

- Incubate overnight at 37°C.
- Prepare and sterilize in the autoclave tubes with 6 ml of liquid LB medium
- Prepare glycerol 40%

3rd Day

Transformation (see **Transformation Protocol in *Escherichia coli* DH5-α**)

Organism: DH5 - α

Selection: Cloranphenicol

4th Day

- Inoculate 3 – 4 colonies in a 6 ml LB with the same antibiotic used in the transformation protocol.
- Incubate overnight at 275rpm/37°C.

5th Day

Miniprep

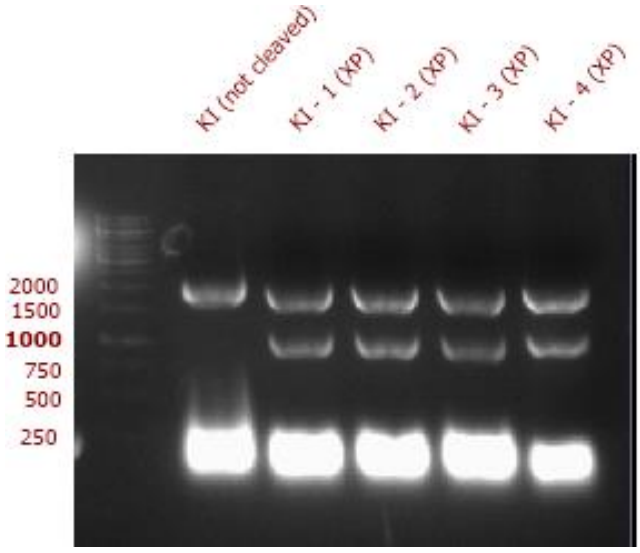
- Prepare **glycerol stock** of the clones (500ul glycerol 40% + 500ul inoculum).
- Extract plasmidial DNA (see **Alkaline Lyses or PureLink Invitrogen Protocol**)
- Run a preliminary electrophoresis gel.
- Quantify DNA samples.

Assembly Confirmation

- XP Digestion (see **Enzymatic Digestion Protocol**)

Assembly	Volume to 300 ng (ul)	Buffer x10 (ul)	XbaI (ul)	PstI (ul)	H_2O to 10ul (ul)
KI – 1	3	1	0,5	0,5	5
KI – 2	3	1	0,5	0,5	5
KI – 3	3	1	0,5	0,5	5
KI – 4	3	1	0,5	0,5	5

- Incubate for 2 hours at 37°C.
- Prepare samples for DNA sequencing.
- Run an electrophoresis analysis of the XP digestion



Size expected	Size in gel
1115 bp	~ 1000 bp